

# 1 Control of Culture Media and Strains of 2 Microorganisms Used for Microbial Test 3 Methods

4 (微生物試験法に用いる培地及び微生物株の管理)

5  
6 This General Information describes points to consider in  
7 the control of culture media and strains of microorganism  
8 used for microbial test methods in a laboratory.

9 Use apparatuses which are appropriately maintained,  
10 controlled, and calibrated.

## 11 1. Media preparation and quality control

### 12 1.1. Media preparation

13 Select culture media and medium components suitable  
14 for microbial tests to be conducted when preparing culture  
15 media. Dehydrated media are accompanied by component  
16 compositions and instructions for preparation. Because dif-  
17 ferent media may have different preparation requirements  
18 (e.g., heating, additives, and pH adjustment), it is important  
19 to follow their instructions to prepare media with appropri-  
20 ate quality. Records of the date of preparation, name, lot  
21 number, mass of dehydrated media or medium components,  
22 the volume of water used, sterilization conditions, pH after  
23 sterilization, and equipment used, etc. are useful to investi-  
24 gate cause when a problem occurs.

25 Dehydrated media or medium components should be  
26 weighed appropriately. In addition, clean containers and  
27 tools should be used to prevent contamination with foreign  
28 matters during preparation. Water suitable for performing  
29 the relevant test should be used to prepare culture media,  
30 and purified water is most often used.

31 Dehydrated media should be dissolved in water before  
32 sterilization, or shaken thoroughly to disperse sufficiently.  
33 When dispensing before sterilization, media should be  
34 thoroughly dissolved in water. If heating is necessary to  
35 dissolve media, care should be taken not to overheat media.  
36 Browning of media by the Maillard reaction etc. is one of  
37 the indication of overheating. Appropriate equipment and  
38 tools should be used for heating, stirring and mixing in the  
39 preparation of media. When adding components that can-  
40 not be heated, they should be aseptically added to media  
41 cooled to an appropriate temperature after sterilization, and  
42 mixed thoroughly.

43 If poorly cleaned tools are used to prepare media, sub-  
44 stances that inhibit the growth of microorganisms may con-  
45 taminate the media. Inhibitory substances are derived from  
46 detergent residues after cleaning tools, etc., substances  
47 used before cleaning tools, or residues during manufactur-  
48 ing even when unused tools are used. In the cleaning pro-

49 cess, residues and foreign matters should be removed cer-  
50 tainly, and finally detergents, etc. should be washed out  
51 completely using purified water, etc.

52 Sterilization of media should be performed within pa-  
53 rameters (temperature, pressure, exposure time, etc.) pro-  
54 vided by a manufacturer or parameters validated by users.  
55 Sterilization in an autoclave is preferred, unless media con-  
56 tain medium components that are unstable to heat. Sterili-  
57 zation by filtration may be appropriate for some medium  
58 compositions.

59 When an autoclave is used, the sterilization should be  
60 performed in the sterilization cycles (which includes the  
61 process where the temperature of products to be sterilized  
62 rises to specified temperature and the process from the  
63 completion of sterilization until the falling to the tempera-  
64 ture to be able to take out the objects) in which the temper-  
65 ature of all products to be sterilized (temperature of media)  
66 meets specified temperature and exposure time, depending  
67 on the loading format of loads (loading pattern: the shape,  
68 size, number and distribution of containers, and the type  
69 and liquid volume of a product to be sterilized, etc.). The  
70 sterilization cycles in which temperature rises slowly may  
71 result in the overheating of media. In general, the more the  
72 liquid volume of an object to be sterilized, the longer the  
73 sterilization cycle. However, since the cycle is affected by  
74 the size and number of dispensing containers even if the  
75 total volume of liquid is the same, so appropriate conditions  
76 should be selected. After the completion of sterilization cy-  
77 cle, the media should be taken out immediately, and cooled,  
78 if necessary. The effects of the sterilization cycles should  
79 be verified by the growth promotion test (refer to Microbi-  
80 ological Examination of Non-sterile Products <4.05>, Steri-  
81 lity Test <4.06>, etc.) together with confirmation of the  
82 sterility of media (no microbial contamination).

83 Take into consideration that improper preparation may  
84 result in the deterioration of growth promoting properties  
85 and inhibitory properties, and in the deviation of the prop-  
86 erties, such as color, clarity, gel strength and pH, from the  
87 acceptable range.

88 The pH of a medium should be confirmed after it has  
89 cooled to room temperature or another specified tempera-  
90 ture by aseptically withdrawing a test sample every each  
91 sterile batch (unit to be sterilized at one time, hereinafter  
92 called as "batch"). If it cannot be measured at the specified  
93 temperature, the pH should be corrected for the specified  
94 temperature. A flat pH probe is recommended for agar sur-  
95 faces, and an immersion probe is recommended for liquids.  
96 The pH of media should be within a specified range. How-  
97 ever, unless when it is confirmed that a wider range is ac-  
98 ceptable by the growth promotion test or suitability test (re-  
99 fer to Microbiological Examination of Non-sterile Products  
100 <4.05>, Sterility Test <4.06>, etc.).

101 Prepared media (agar plate or media dispensed to test  
102 tubes etc.) are identified by name, batch number, prepara-  
103 tion date, etc. Also, pay attention to the following terms.

- 104 (i) Container fracture
- 105 (ii) Unequal dispensing volume between containers
- 106 (iii) Dirt of containers due to adhesion of medium  
107 components, etc.
- 108 (iv) Browning or discoloration
- 109 (v) Air bubbles
- 110 (vi) Status of redox indicator (if applicable)
- 111 (vii) Hemolysis (if applicable)
- 112 (viii) Formation of crystals, etc.
- 113 (ix) Drying that causes cracks and dimples
- 114 (x) Microbial contamination

### 115 1.2. Media storage

116 When storing media and medium components, the fol-  
117 lowing points should be noted, including transport condi-  
118 tions until acquisition, in order to prevent the deterioration  
119 of quality.

- 120 (i) Drying, evaporation, moisture absorption
- 121 (ii) Temperature
- 122 (iii) Microbial contamination
- 123 (iv) Contamination of foreign matters
- 124 (v) Fracture

125 In addition, media or medium components should be la-  
126 belled with names, batch or lot numbers, storage conditions,  
127 expiration dates, etc. and identified.

128 The storage conditions and the expiration dates of the  
129 media after preparation is set after the stability is confirmed  
130 by verifying that the performance of the media meet the ac-  
131 ceptance criteria up to the end of the expiration date by the  
132 growth promotion test and other necessary quality tests  
133 when media are stored under the set conditions.

134 For long-term storage, packaging materials, packaging  
135 types, containers and stoppers that can prevent evaporation  
136 of water should be selected. Also, protect from light, if nec-  
137 essary. The agar media should be stored avoiding freezing  
138 because freezing damage the gel structure of agar.

139 In addition, agar media that have been remelted after  
140 storage can be used within the confirmed expiration date, if  
141 performance tests are performed to confirm the suitability.  
142 Also, it is desirable to remelt agar media only once to avoid  
143 the possibility of deterioration and contamination due to  
144 overheating. It is recommended that remelting is performed  
145 in a heated water bath or in free-flowing steam. When using  
146 a microwave oven or a heating plate for melting media, care  
147 should be taken because whole media may not be uniformly  
148 heated, and deterioration of the media and breakage of the  
149 container due to overheating may occur.

150 Agar media immediately after sterilization, or remelted  
151 media should be held at 45 to 50°C or another specified  
152 temperature, but holding for a long-time should be avoided

153 in consideration of the risk of deterioration and contamina-  
154 tion. In addition, if media are held in a water bath, be care-  
155 ful of contamination derived from water in a bath when  
156 pouring them into petri dishes.

157 When discarding used or expired media, sterilize as  
158 needed and take care to prevent contamination.

### 159 1.3. Quality control testing

160 Perform the following quality control tests for each batch  
161 or lot for all prepared media. The prepared media include  
162 ready-prepared media and ready-to use swabs, strips, etc.

- 163 (i) Growth promotion (growth promoting properties,  
164 and, as needed, inhibitory properties or indicative proper-  
165 ties)
- 166 (ii) pH (as needed, for ready-prepared media)

167 Media purchased or stored under a refrigeration condi-  
168 tion should be returned to room temperature or a tempera-  
169 ture being specified separately, and confirmed.

- 170 (iii) Sterility (no microbial contamination)

171 Media used for environmental monitoring of Grade A  
172 and B in processing areas for sterile pharmaceutical prod-  
173 ucts should be multiple-wrapped or carried into the areas  
174 according to a specified procedure after disinfecting or de-  
175 contaminating the exterior. If sterilization after packaging  
176 is not performed, all media (100%) should be subjected to  
177 incubation prior to use and confirmed to be free of micro-  
178 bial contamination in order to prevent extraneous contami-  
179 nation from being carried into controlled environments and  
180 prevent false-positive results.

181 A certificate of analysis describing storage conditions  
182 and an expiration date accompanies ready-prepared media,  
183 as well as the standard microbial strain used in growth pro-  
184 motion testing. When the data of testing are obtained from  
185 a media manufacturer at the time of acceptance, if the data  
186 is reliable as the result of writing or site investigation etc.  
187 and the validity of the growth promotion testing result and  
188 the expiration date can be confirmed, the quality control  
189 test may be performed regularly instead of every batch or  
190 lot.

191 Similar to media, those requiring quality control tests in-  
192 clude reagents for microbial identification tests such as  
193 Gram staining reagents and oxidase reagents. These should  
194 be subjected to quality control tests at the time of ac-  
195 ceptance or use using appropriate standard microbial  
196 strains selected.

### 197 2. Maintenance and control of microbial strains

198 The appropriate treatment of stock microbial strains is  
199 very important to maintain the accuracy and repeatability  
200 of results of microbial tests. The storage and handling of  
201 microbial strains in a laboratory should be done in such a  
202 way that will minimize changes in the growth characteris-  
203 tics of the microorganism, paying attention to contamina-  
204 tion. Microbial strains used in the compendial methods are

205 available in frozen, lyophilized, slant cultured or ready-to-  
206 use forms from an organization for culture collections or an  
207 appropriate supplier. Obtained microbial strains should be  
208 confirmed to be contaminated with no other strains before  
209 or at use for quality control testing by the observation of  
210 emerged colonies being single, etc. when spread on plate  
211 media having no selectivity. In addition, if necessary, con-  
212 firm the distributed microbial species.

213 Stock microbial strains are resuscitated according to the  
214 method specified by an organization for culture collections,  
215 etc. The application of seed lot culture maintenance tech-  
216 niques (seed-lot systems) is recommended to control the  
217 preservation of microbial strains. The seed-lot system is a  
218 system to control the number of passage of stock microbial  
219 strains in order to avoid property changes due to passages.  
220 One passage is defined as a transfer to a fresh medium. Any  
221 form of subculturing is considered to be a passage. At least  
222 the following terms should be noted in the control.

223 (i) Count a culture obtained by resuscitating (the num-  
224 ber of passage is first) a microbial strain obtained from an  
225 organization for culture collections, etc. as the first genera-  
226 tion.

227 (ii) Control the number of passages.

228 (iii) Microbial strains must not be used more than 5  
229 passages for growth promotion tests and suitability tests.

230 There are methods by freezing, by drying, by serial sub-  
231 culture, etc. for the preservation of microbial strains, and  
232 an example of the method by freezing is shown. A standard  
233 microbial strain obtained from an organization for culture  
234 collections etc. is resuscitated and grown in appropriate  
235 medium. Aliquot of this culture (the first generation) is sus-  
236 pended in a solution containing a protective agent that pre-  
237 vents freezing damage, transferred to a vial or the like, and  
238 cryopreserved at an appropriate temperature according to  
239 the microbial species. Many microbial strains can be stored  
240 for a long time by maintaining at the temperature not ex-  
241 ceeding  $-70^{\circ}\text{C}$ . If the second and subsequent generations  
242 are prepared in large quantities, the frequency of obtaining  
243 and preparing standard microbial strains can be reduced.

244 Microbial strains in once opened containers should be  
245 discarded without refreezing to avoid the risk of reduced  
246 viability and contamination of stored microbial strains.

## 247 **References**

- 248 1) WHO, WHO good practices for pharmaceutical micro-  
249 biology laboratories(WHO Technical Report Series,  
250 No.961, Annex 2, 2011).
- 251 2) US Pharmacopeia 41 (2018), <1117> Microbiological  
252 Best Laboratory Practices.

253

254