

## 1 Bacterial Endotoxins Test and alternative methods using recombinant protein-reagents for endotoxin assay

(エンドトキシン試験法と測定試薬に遺伝子組換えタンパク質を用いる代替法)

Endotoxins, also called lipopolysaccharides, are present in the outer cell membrane of Gram-negative bacteria and exhibit various biological activities. Endotoxins, when entering the blood stream, can cause fever even in a very small quantity, and a large quantity of endotoxins is very toxic and can cause death due to endotoxin shock. In addition, endotoxins may contaminate pharmaceutical preparations during the production process because these are derived from Gram-negative bacteria widely present in the environment and because these are hard to be inactivated due to their heat-resistance. Endotoxins are designated as substances which should be controlled to ensure the safety of pharmaceutical preparations, etc., because these exhibit higher pyrogenicity than other well-known pyrogens which may contaminate them. Bacterial Endotoxins Test <4.01> is an *in vitro* test method that can detect endotoxins with high sensitivity using amoebocyte lysate prepared from blood corpuscle extracts of horseshoe crabs, and is applicable to injections, etc. On the other hand, recombinant protein-reagents for endotoxin assay have been developed as alternatives to lysate reagents for the purpose of protecting horseshoe crabs, ensuring a stable supply of reagents, reducing differences between reagent lots, and improving the continuity of the tests.

This General Information describes procedures and consideration in measurement when using recombinant protein-reagents for endotoxin assay as alternative methods, in addition to lysate reagents and test methods in Bacterial Endotoxins Test <4.01>.

### 1. Measurement principle of the Bacterial Endotoxins Test

Bacterial Endotoxins Test <4.01> is a test to detect or quantify bacterial endotoxins using amoebocyte lysate prepared from blood corpuscle extracts of horseshoe crab (*Limulus polyphemus* or *Tachypleus tridentatus*). This test utilizes the reaction in which the hemocyte extract of horseshoe crab is coagulated by endotoxins, and the coagulation reaction is based on a chain reaction by multiple serine proteases triggered by endotoxins (Fig. 1). Endotoxins activate factor C contained in the hemocyte extract of horseshoe crab to convert to an active serine protease, which in turn successively activates factor B, and then proclotting enzyme. Finally, coagulogen, which is a coagulant protein, is hydrolyzed to result in coagulin, and insoluble gel is

formed and solidified. In addition, the hemocyte extract of horseshoe crab reacts not only to endotoxins but also to  $\beta$ -glucans and coagulates by a chain reaction starting from factor G.

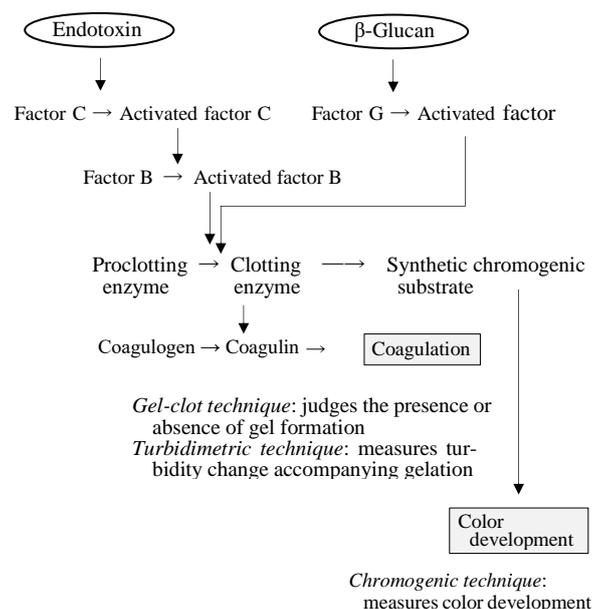
### 2. Measurement method in the Bacterial Endotoxins Tests

Bacterial Endotoxins Test <4.01> includes the gel-clot techniques, which are based on the gel formation of the lysate TS, and the photometric quantitative techniques, which are based on endotoxin-induced optical changes (Fig. 1).

The gel-clot techniques visually confirm the presence or absence of gel formation and require no special device for the determination. The gel-clot techniques include a limit test and a quantitative test. The former is a method for judging whether a sample contains endotoxins exceeding the endotoxin limit specified in each monograph, using the labeled sensitivity of a lysate reagent as an index. The latter is a method for quantifying the amount of endotoxins in a sample by determining an endpoint, which is defined as the maximum dilution of a sample solution showing the gel formation.

The photometric quantitative techniques include the turbidimetric technique and the chromogenic technique (Fig. 1). In both techniques, the lysate TS and a sample solution are mixed, and the reaction solution is measured after a given time or over time using a spectrophotometer. The turbidimetric technique measures turbidity changes accompanying gelation of the lysate TS using absorbance or transmittance, and the chromogenic technique measures the amount of chromophore released from a synthetic chromogenic substrate by the reaction of endotoxins with the lysate TS using absorbance or transmittance.

83



84 **Fig. 1** Measurement principle and method in endotoxin  
85 assay

### 86 3. Reagents used for the Bacterial Endotoxins Test

87 There are several lysate reagents used for the Bacterial  
88 Endotoxins Test<4.01> corresponding to each test method.  
89 The reagents are classified into two types based on their  
90 reactivity to endotoxins and  $\beta$ -glucans. One is a reagent  
91 which contains both the reaction system starting from fac-  
92 tor C and the reaction system starting from factor G. The  
93 other is a reagent which detects only endotoxins by the re-  
94 action system starting from factor because it does not con-  
95 tain factor G or suppresses the reaction of factor G system.  
96 Appropriate reagents should be selected depending on the  
97 sample to be examined and the purpose of the test.

98 On lysate reagents used for the gel-clot techniques, the  
99 lowest concentration of endotoxins that cause coagulation  
100 (gel formation) is set and labeled as the labeled lysate rea-  
101 gent sensitivity (endotoxin unit (EU)/mL) by the reagent  
102 manufacturers. The acceptance of a sample is judged using  
103 the labeled sensitivity as an index. In order to obtain accu-  
104 rate test results, confirm that the labeled sensitivity is ap-  
105 propriate according to 4.1.1. Test for confirmation of la-  
106 beled lysate reagent sensitivity in Bacterial Endotoxins  
107 Test <4.01> 4.1. Preparatory testing. If the geometric mean  
108 endpoint concentration does not fall within the specified  
109 range, repeat the test after adjusting test conditions. If the  
110 geometric mean endpoint concentration does not fall within  
111 the specified range by the retest, the lysate reagent cannot  
112 be used.

113 When using lysate reagents for the photometric quanti-  
114 tative techniques, for both turbidimetric and chromogenic  
115 techniques, a standard curve is prepared using the standard  
116 solutions of three or more concentrations within the quan-  
117 titative concentration range. Confirm that the test proce-  
118 dures of an operator and test conditions are appropriate ac-  
119 cording to 5.3.1. Test for assurance of criteria for the stand-  
120 ard curve in the Bacterial Endotoxins Test <4.01> 5.3. Pre-  
121 paratory testing. Although the labeled sensitivity is not  
122 shown on the lysate reagents used for the photometric  
123 quantitative techniques, the lowest concentration of the  
124 standard solution used for the generation of the standard  
125 curve corresponds to the labeled sensitivity.

126 Most pharmaceuticals are found to interfere with the  
127 Bacterial Endotoxins Tests <4.01> performance, although  
128 to greater or lesser degrees. In general, the influence of in-  
129 terfering factors present in a sample solution can often be  
130 overcome by dilution. In this case, samples should be di-  
131 luted with water for bacterial endotoxins test within the  
132 range that does not exceed a maximum valid dilution for  
133 the measurement. The Maximum Valid Dilution is the  
134 maximum allowable dilution of a sample solution. As

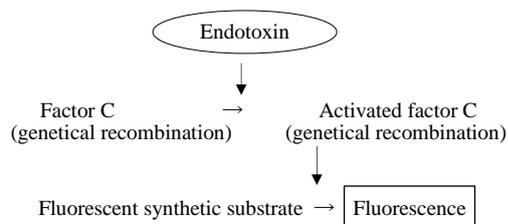
135 shown in Bacterial Endotoxins Test <4.01> 3. Determina-  
136 tion of Maximum Valid Dilution,  $\lambda$  is a labeled sensitivity  
137 for a lysate reagent in the gel-clot techniques and is the low-  
138 est concentration of a standard curve for a lysate reagent in  
139 the photometric quantitative techniques; the smaller the  $\lambda$ ,  
140 the larger the maximum valid dilution.  $\lambda$  for many lysate  
141 reagents used for the photometric quantitative techniques is  
142 smaller than those for lysate reagents used for the gel-clot  
143 techniques. If interfering factors contained in a sample are  
144 definite, perform procedures to reduce them. If interfering  
145 factors cannot be reduced or interference cannot be avoided  
146 because of indefinite interfering factors, consider using  
147 other lysate reagents or changing the test method.

### 148 4. Measurement by alternative methods using recom- 149 binant protein-reagents for endotoxin assay and points 150 to consider in the measurement

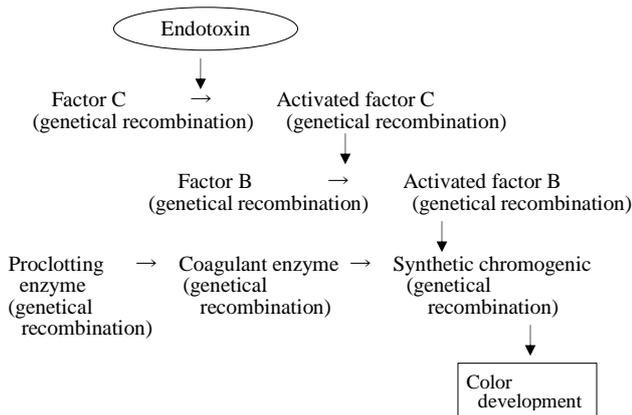
151 Recombinant protein-reagents for endotoxin assay are  
152 classified into two types, namely both reagents are com-  
153 posed of the recombinant protein(s) derived from blood  
154 corpuscle extracts of horseshoe crabs. The first are reagents  
155 mainly composed of the recombinant protein of factor C  
156 (horseshoe crab: *Carcinoscorpius rotundicauda* or *T. tri-*  
157 *dentatus*), and the second are reagents composed of the re-  
158 combinant proteins of factor C, factor B and proclotting en-  
159 zyme (horseshoe crab: *T. tridentatus*). The former reagents  
160 measure the amount of fluorescence generated by cleavage  
161 of a fluorescent synthetic substrate by activated factor C  
162 (genetical recombination) which has been activated by en-  
163 dotoxins (Fig. 2a). In the case of the latter reagents com-  
164 posed of the three kinds of recombinant proteins, endotox-  
165 ins activate factor C (genetical recombinant), and it further  
166 activates factor B (genetical recombinant) and proclotting  
167 enzyme successively in the same way as the measurement  
168 method of the chromogenic technique. Then, the amount of  
169 chromophore released from the synthetic chromogenic sub-  
170 strate is measured (Fig. 2b). In the both cases, a reagent so-  
171 lution and a sample solution are mixed, keeping at  $37 \pm 1^\circ\text{C}$ ,  
172 and the fluorescence intensity or absorbance of the reaction  
173 solution is measured optically after a given time or over  
174 time.

175 The recombinant protein-reagents for endotoxin assay  
176 do not correspond to "an amoebocyte lysate prepared from  
177 blood corpuscle extracts of horseshoe crab" specified in  
178 Bacterial Endotoxins Test <4.01>. If these reagents for en-  
179 dotoxin assay are used as an alternative method, confirm  
180 that accuracy, precision, sensitivity, specificity, etc. are  
181 equal or better compared to Bacterial Endotoxins Test  
182 <4.01> using the lysate TS. Among the recombinant pro-  
183 tein-reagents for endotoxin assay, some have been reported  
184 to have sensitivity and specificity equal to or better than the  
185 method using the lysate TS. When bacterial endotoxins  
186 tests <4.01> for pharmaceuticals, etc. are performed using

187 recombinant protein-reagents, it is necessary to conduct  
 188 5.3.1. Test for assurance of criteria for the standard curve as  
 189 with the Photometric quantitative techniques in the Bacte-  
 190 rial Endotoxins Test <4.01>, and in that case the lowest con-  
 191 centration of the standard curve corresponds to  $\lambda$  (EU/mL).  
 192 In addition, it is necessary to pay attention to interference,  
 193 and Test for interfering factors should be performed. Particu-  
 194 larly, in the case of methods that measure the amount  
 195 of fluorescence, it is necessary to be careful of interference  
 196 because even substances that do not disturb the measure-  
 197 ment when using lysate reagents may exhibit interference  
 198 such as the inhibition of the generation of fluorescence. In  
 199 addition, regarding reagents prepared using the gene se-  
 200 quence of factor C of different kinds of horseshoe crab from  
 201 horseshoe crab (*L. polyphemus* or *T. tridentatus*) specified  
 202 in the Bacterial Endotoxins Test, it should be noted that the  
 203 difference of factor C may affect the reactivity to endotox-  
 204 ins.  
 205



206 **Fig. 2a** Reagents for endotoxin assay mainly composed  
 207 of recombinant factor C  
 208



209 **Fig. 2b** Reagents for endotoxin assay composed of the  
 210 three kinds of recombinant proteins  
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