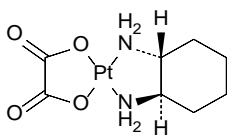


1 Oxaliplatin

2 オキサリプラチン



4 $C_8H_{14}N_2O_4Pt$: 397.29

5 (SP-4-2)-[(1R,2R)-Cyclohexane-1,2-diamine- $\kappa N, \kappa N'$][ethanedioato

6 (2-)- $\kappa O^1, \kappa O^2$]platinum

7 [61825-94-3]

8

9 Oxaliplatin contains not less than 98.0% and not
10 more than 102.0% of oxaliplatin ($C_8H_{14}N_2O_4Pt$), calcu-
11 lated on the dried basis.

12 **Description** Oxaliplatin occurs as a white crystalline pow-
13 der.

14 It is slightly soluble in water, very slightly soluble in meth-
15 anol, and practically insoluble in ethanol (99.5).

16 Optical rotation $[\alpha]_D^{20}$: +74.5 – +78.0° (0.25 g calcu-
17 lated on the dried basis, water, 50 mL, 100 mm).

18 **Identification (1)** To 2 mL of a solution of oxaliplatin (1
19 in 500) add 2 to 3 drops of diluted tin (II) chloride TS (1 in
20 15), and allow to stand for 30 minutes: a yellow to orange-
21 yellow precipitate is formed.

22 **(2)** Determine the absorption spectrum of a solution of
23 Oxaliplatin (1 in 10,000) as directed under Ultraviolet-visible
24 Spectrophotometry <2.24>, and compare the spectrum with
25 the Reference Spectrum or the spectrum of a solution of Ox-
26 aliplatin RS prepared in the same manner as the sample solu-
27 tion: both spectra exhibit similar intensities of absorption at
28 the same wavelengths.

29 **(3)** Determine the infrared absorption spectrum of Oxali-
30 platin as directed in the potassium bromide disk method un-
31 der Infrared Spectrophotometry <2.25>, and compare the
32 spectrum with the Reference Spectrum or the spectrum of
33 Oxaliplatin RS: both spectra exhibit similar intensities of ab-
34 sorption at the same wave numbers.

35 **Purity (1)** Acidity or Alkalinity—Dissolve 0.20 g of Ox-
36 aliplatin in freshly boiled and cooled water to make 100 mL.
37 To 50 mL of this solution add 0.5 mL of phenolphthalein TS:
38 no color develops. To this solution add 0.6 mL of 0.01 mol/L
39 sodium hydroxide TS: a pale red color develops.

40 **(2)** Oxalic acid—Conduct this procedure within 20
41 minutes after preparation of the sample solution. Dissolve ex-
42 actly 0.100 g of Oxaliplatin in water to make exactly 50 mL,
43 and use this solution as the sample solution. Separately, dis-
44 solve exactly 14 mg of oxalic acid dihydrate in water to make

45 exactly 250 mL. Pipet 5 mL of this solution, add water to
46 make exactly 100 mL, and use this solution as the standard
47 solution. Perform the test with exactly 20 μL each of the sam-
48 ple solution and standard solution as directed under Liquid
49 Chromatography <2.01> according to the following condi-
50 tions. Determine the peak areas of oxalic acid in each solution
51 by the automatic integration method: the peak area of oxalic
52 acid obtained from the sample solution is not larger than the
53 peak area of oxalic acid from the standard solution.

54 **Operating conditions—**

55 **Detector:** An ultraviolet absorption photometer (wave-
56 length: 205 nm).

57 **Column:** A stainless steel column 4.6 mm in inside diam-
58 eter and 25 cm in length, packed with octadecylsilanized sil-
59 ica gel for liquid chromatography (5 μm in particle diameter).

60 **Column temperature:** A constant temperature of about
61 40°C.

62 **Mobile phase:** Dissolve 2.6 mL of 40% tetrabutylammo-
63 nium hydroxide TS and 1.36 g of potassium dihydrogen
64 phosphate in water to make 1000 mL, and adjust to pH 6.0
65 with phosphoric acid. To 800 mL of this solution add 200 mL
66 of acetonitrile for liquid chromatography.

67 **Flow rate:** 2.0 mL per minute.

68 **System suitability—**

69 **System performance:** When the procedure is run with 20
70 μL of the standard solution under the above operating condi-
71 tions, the number of theoretical plates and the symmetry fac-
72 tor of the peak of oxalic acid is not less than 5000 and not
73 more than 2.0, respectively.

74 **System repeatability:** When the test is repeated 6 times
75 with 20 μL of the standard solution under the above operating
76 conditions, the relative standard deviation of the peak area of
77 oxalic acid is not more than 3.0%.

78 **(3) Related substance A—**Conduct this procedure within
79 20 minutes after preparation of the sample solution. Weigh
80 accurately about 0.1 g of Oxaliplatin, dissolve in water to
81 make exactly 50 mL, and use this solution as the sample so-
82 lution. Separately, weigh accurately about 12.5 mg of Oxali-
83 platin Related Substance A Dinitrate for Purity RS, dissolve
84 in about 63 mL of methanol, and add water to make exactly
85 250 mL. Pipet 5 mL of this solution add water to make ex-
86 actly 100 mL, and use this solution as the standard solution.
87 Perform the test with exactly 20 μL each of the sample solu-
88 tion and standard solution as directed under Liquid Chroma-
89 tography <2.01> according to the following conditions. De-
90 termine the peak areas, A_{T1} and A_S , of the related substance A
91 in the sample solution and standard solution by the automatic
92 integration method, and calculate the amount of the related
93 substance A by the following equation: the amount of the re-
94 lated substances A is not more than 0.1%.

95
$$\text{Amount (\%)} \text{ of the related substance A}$$

96
$$= M_S / M_T \times A_{T1} / A_S \times 0.797$$

97 M_S : Amount (mg) of Oxaliplatin Related Substance A
98 Dinitrate for Purity RS taken
99 M_T : Amount (mg) of Oxaliplatin taken
100 0.797: Conversion factor for the related substance A dinitrate to related substance A
101

102 *Operating conditions—*

103 Detector: An ultraviolet absorption photometer (wavelength: 215 nm).

105 Column: A stainless steel column 4.6 mm in inside diameter and 25 cm in length, packed with octadecylsilanized silica gel for liquid chromatography (5 μ m in particle diameter).

108 Column temperature: A constant temperature of about 109 40°C.

110 Mobile phase: Dissolve 1.36 g of potassium dihydrogen phosphate and 1 g of sodium 1-heptane sulfonate in 1000 mL of water, and adjust to pH 3.0 with phosphoric acid. To 800 mL of this solution add 200 mL of acetonitrile for liquid chromatography.

115 Flow rate: 2.0 mL per minute.

116 Time span of measurement: About 2.5 times as long as the retention time of the related substance A, beginning after the solvent peak.

119 *System suitability—*

120 Test for required detectability: Pipet 1 mL of the standard solution, add water to make exactly 10 mL. Confirm that the peak area of the related substance A obtained with 20 μ L of this solution is equivalent to 7 to 13% of that with 20 μ L of the standard solution.

125 System performance: Heat a solution of oxaliplatin in diluted dilute sodium hydroxide TS (1 in 20) (1 in 500) at 60°C for about 2 hours, and allow to cool. To 1 mL of this solution add water to make exactly 10 mL. When the procedure is run with 20 μ L of this solution under the above operating conditions, the resolution between the related substance A and the peak having the relative retention time of about 1.4 to the related substance A is not less than 4, and the symmetry factor of the peak of the related substance A is not more than 2.0.

134 System repeatability: When the test is repeated 6 times with 20 μ L of the standard solution under the above operating conditions, the relative standard deviation of the peak area of the related substance A is not more than 3.0%.

138 (4) Other related substances—Conduct this procedure within 20 minutes after preparation of the sample solution. Dissolve 0.10 g of Oxaliplatin in water to make 50 mL, and use this solution as the sample solution. Pipet 1 mL of the sample solution, and add water to make exactly 100 mL. Then, pipet 5 mL of this solution, and add water to make exactly 50 mL, and use this solution as the standard solution. Perform the test with exactly 10 μ L each of the sample solution and standard solution as directed under Liquid Chromatography <2.01> according to the following conditions, and determine each peak area by the automatic integration

149 method: the peak area of the related substance B, having the relative retention time of about 0.6 to oxaliplatin, obtained from the sample solution is not larger than 4.4 times the peak area of oxaliplatin from the standard solution. Furthermore, the total area of the peaks other than oxaliplatin and the peak mentioned above from the sample solution is not larger than the peak area of oxaliplatin from the standard solution.

156 *Operating conditions—*

157 Detector, column, column temperature, mobile phase, and flow rate: Proceed as directed in the Assay.

159 Time span of measurement: About 3 times as long as the retention time of oxaliplatin, beginning after the solvent peak.

161 *System suitability—*

162 Test for required detectability: Pipet 1 mL of the standard solution, and add water to make exactly 10 mL. Confirm that the peak area of oxaliplatin obtained with 20 μ L of this solution is equivalent to 7 to 13% of that with 20 μ L of the standard solution.

167 System performance: To 1 mL of a solution of oxaliplatin (1 in 500) and 1 mL of 1 mol/L sodium chloride TS add water to make 10 mL. Separately, to 1 mL of a solution of oxaliplatin (1 in 500) and 1 mL of diluted hydrogen peroxide (30) (1 in 3000) add water to make 10 mL. Heat these solutions at 60°C for about 2 hours, and allow to cool. Mix 1 mL each of these solutions, and add water to make 10 mL. When the procedure is run with 10 μ L of this solution under the above operating conditions, the resolution between the peak having the relative retention time of about 0.9 to oxaliplatin and oxaliplatin is not less than 2.0, and the symmetry factor of oxaliplatin is not less than 2.0.

179 System repeatability: When the test is repeated 6 times with 10 μ L of the standard solution under the above operating conditions, the relative standard deviation of the peak area of oxaliplatin is not more than 3.0%.

183 (5) Optical isomer—Dissolve 30 mg of Oxaliplatin in methanol to make 50 mL, and use this solution as the sample solution. Pipet 5 mL of the sample solution, and add methanol to make exactly 100 mL. Pipet 2 mL of this solution, add methanol to make exactly 100 mL, and use this solution as the standard solution. Perform the test with exactly 20 μ L each of the sample solution and standard solution as directed under Liquid Chromatography <2.01> according to the following conditions. Determine each peak height in each solution by the automatic peak height method: the height of the peak having the relative retention time of about 1.2 to oxaliplatin obtained from the sample solution is not higher than the peak height of oxaliplatin from the standard solution.

196 *Operating conditions—*

197 Detector: An ultraviolet absorption photometer (wavelength: 254 nm).

199 Column: A stainless steel column 4.6 mm in inside diameter and 25 cm in length, packed with silica gel coated with

201 phenylcarbamoyl cellulose for liquid chromatography (5 μm
202 in particle diameter).

203 Column temperature: A constant temperature of about
204 40°C.

205 Mobile phase: A mixture of methanol and ethanol (99.5)
206 (7:3).

207 Flow rate: 0.3 mL per minute.

208 *System suitability*—

209 System performance: When the procedure is run with 20
210 μL of the standard solution under the above operating condi-
211 tions, the number of theoretical plates and the symmetry fac-
212 tor of the peak of oxaliplatin are not less than 5000 and not
213 more than 2.0, respectively.

214 System repeatability: When the test is repeated 6 times
215 with 20 μL of the standard solution under the above operating
216 conditions, the relative standard deviation of the peak area of
217 oxaliplatin is not more than 3.0%.

218 **Loss on drying** <2.41> Not more than 0.5% (1 g, 105°C, 2
219 hours).

220 **Assay** Weigh accurately about 20 mg each of Oxaliplatin
221 and Oxaliplatin RS (separately determine the loss on drying
222 <2.41> under the same conditions as Oxaliplatin), dissolve
223 each in water to make exactly 200 mL, and use these solu-
224 tions as the sample solution and the standard solution, respec-
225 tively. Perform the test with exactly 20 μL each of the sample
226 solution and standard solution as directed under Liquid Chro-
227 matography <2.01> according to the following conditions,
228 and determine the peak areas, A_T and A_S , of oxaliplatin in
229 each solution.

230 Amount (mg) of oxaliplatin ($\text{C}_8\text{H}_{14}\text{N}_2\text{O}_4\text{Pt}$)
231 $=M_S \times A_T/A_S$

232 M_S : Amount (mg) of Oxaliplatin RS taken, calculated on
233 the dried basis

234 *Operating conditions*—

235 Detector: An ultraviolet absorption photometer (wave-
236 length: 210 nm).

237 Column: A stainless steel column 4.6 mm in inside diam-
238 eter and 25 cm in length, packed with octadecylsilanized sil-
239 ica gel for liquid chromatography (5 μm in particle diameter).

240 Column temperature: A constant temperature of about
241 40°C.

242 Mobile phase: Adjust the pH of 1000 mL of water to 3.0
243 with phosphoric acid. To 990 mL of this solution add 10 mL
244 of acetonitrile for liquid chromatography.

245 Flow rate: 1.2 mL per minute.

246 *System suitability*—

247 System performance: When the procedure is run with 20
248 μL of the standard solution under the above operating condi-
249 tions, the number of theoretical plates and the symmetry

250 factor of the peak of oxaliplatin are not less than 3000 and
251 not more than 2.0, respectively.

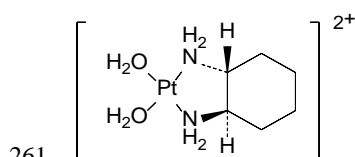
252 System repeatability: When the test is repeated 6 times
253 with 20 μL of the standard solution under the above operating
254 conditions, the relative standard deviation of the peak area of
255 oxaliplatin is not more than 1.0%.

256 **Containers and storage** Containers—Tight containers.

257 **Others**

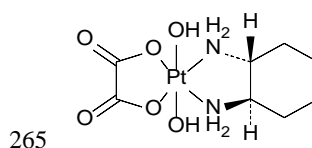
258 Related substance A:

259 (*SP-4-2*)-Diaqua[(1*R*,2*R*)-cyclohexane-1,2-diamine- κN , $\kappa\text{N}'$]
260 platinum



262 Related substance B:

263 (*OC-6-33*)-[(1*R*,2*R*)-Cyclohexane-1,2-diamine- κN , $\kappa\text{N}'$]
264 [ethanedioato(2-)- κO^1 , κO^2] dihydroxyplatinum



266 **Add the following to 9.01 Reference**
267 **Standards (1):**

268 Oxaliplatin RS

269 Oxaliplatin Related Substance A Dinitrate for Purity RS

270 **Add the following to 9.42 Solid Sup-**
271 **ports/Column Packings for Chromatog-**
272 **raphy:**

273 **Silica gel coated with phenylcarbamylated cellulose for**
274 **liquid chromatography** Prepared for liquid chromatog-
275 raphy.