

## 1 **Points to Consider in Suitability Testing, etc.** 2 **of Microbial Tests <G4-12-190>**

3 (微生物学的試験法の適合性試験等における留意事項  
4 <G4-12-190>)

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6 This general information provides points to consider in the  
7 suitability tests of Microbiological Examination of Non-sterile  
8 Products <4.05>, Sterility Test <4.06>, Microbial Limit  
9 Test for Crude Drugs and Preparations containing Crude  
10 Drugs as Main Ingredient <5.02> and Preservatives-Effectiveness  
11 Tests <G4-3-170>.

### 12 **1. Performing tests**

13 Suitability testing of microbial test methods is preferable  
14 to be conducted before or simultaneously with the first mi-  
15 crobiological test on the product to be examined to confirm  
16 the ability of a chosen test method to detect micro-organisms.  
17 If there are any changes in the test method or the product for-  
18 mulation that may affect the test results, the suitability should  
19 be confirmed again.

### 20 **2. Selection and preparation of test strains**

21 Micro-organisms have a variety of natures, which affect  
22 their responses to antimicrobial activity and neutralizing  
23 methods required for their recovery. Depending on the pur-  
24 pose of a test, representative test strains are selected from  
25 bacteria, yeasts, and moulds.

26 The growth state and the preparation method of the test  
27 strains affect the physiological activity of the cells of the test  
28 strains. The use of standardized and stable suspensions or the  
29 use of a freshly prepared inoculum prepared on a specified  
30 liquid medium or agar medium allow for homogeneous and  
31 less variable preparation or reproducible preparation. In Pre-  
32 servatives-Effectiveness Tests <G4-3-170>, it is preferable to  
33 use a freshly prepared inoculum because the bioactivity of  
34 the test strains is likely to affect the results.

### 35 **3. Preparation of test samples**

36 If the preparation processes of test samples may affect mi-  
37 cro-organisms present in the product to be examined, confirm  
38 the effects of the preparation processes. If the sample prepa-  
39 ration is allowed to stand before testing, confirm that there is  
40 no change in the number of micro-organisms, etc.

### 41 **4. General additional procedures for test samples that** 42 **have antimicrobial activity**

43 When antimicrobial activity is observed in the product to  
44 be examined and the test method is changed to remove the  
45 antimicrobial activity, repeat the suitability test using the  
46 changed test method to verify the reproducibility of the re-  
47 sults.

#### 48 **4.1. Dilution**

49 Neutralize the antimicrobial activity of the product to be  
50 examined by dilution. The effective dilution factor varies de-  
51 pending on the product to be examined. If the dilution factor  
52 required for neutralization is too large, neutralize the anti-  
53 microbial activity by other methods.

54 If dilution makes it difficult to interpret results based on  
55 the microbial acceptance criterion in Microbial Attributes of  
56 Non-sterile Pharmaceutical Products <G4-1-170> specified in  
57 the product testing, the number of plates or media may be  
58 increased as appropriate.

#### 59 **4.2. Use of neutralizing agents**

60 For the use of neutralizing agents, refer to 3.4.3. of "I. Mi-  
61 crobiological Examination of Non-sterile products: Micro-  
62 bial Enumeration Tests" in Microbiological Examination of  
63 Non-sterile Products <4.05>. For common neutralizing agents  
64 for interfering substances, refer to Table 4.05-I-2 in Microbi-  
65 ological Examination of Non-sterile Products <4.05>.

66 If the product to be examined is an antibiotic, it may not  
67 be susceptible to neutralization by general neutralizing  
68 agents, but rather by enzymatic treatments (penicillinase,  $\beta$ -  
69 lactamase, etc.). In the case of enzymatic treatment, an ap-  
70 propriate concentration referring to the information such as  
71 Minimum Inhibitory Concentration (MIC) is preferred.

#### 72 **4.3. Membrane filtration**

73 Generally, membrane filters are used. Membrane filters  
74 are made of various materials, so select appropriate material  
75 by considering the properties and composition of the product  
76 to be examined. For example, cellulose mixed ester filters are  
77 used for test samples without antimicrobial activity, cellulose  
78 nitrate filters are used for aqueous, oily, or weakly alcoholic  
79 solutions, and cellulose acetate filters are used for strongly  
80 alcoholic solutions. Low binding filters such as polyvinyl-  
81 idene fluoride (PVDF) are also used to reduce the binding of  
82 antimicrobial substances on the filter.

83 Filters can be washed with an appropriate solution to re-  
84 move antimicrobial substances from the product to be exam-  
85 ined. If the suitability is verified, a surfactant such as poly-  
86 sorbate 80 (concentration: 1 g/L) may be added to the wash-  
87 ings to neutralize residual or bound antimicrobial substances  
88 on the filter or to suspend the test sample.

### 89 **5. Inoculation of test strains**

90 In the microbial enumeration tests of Microbiological Ex-  
91 amination of Non-sterile Products <4.05> and Microbial  
92 Limit Test for Crude Drugs and Preparations containing  
93 Crude Drugs as Main Ingredient <5.02>, when a test sample  
94 is prepared, the initial prepared test sample (lowest dilution)  
95 should be inoculated with test strains to confirm the effect of  
96 antimicrobial activity. Even if the test sample is further di-  
97 luted and tested, inoculate the initial test sample with test  
98 strains. Even when a diluent containing a neutralizing agent  
99 is used, inoculate the initial test sample with test strains. If  
100 antimicrobial activity cannot be removed or neutralized by

101 various means, such as dilution, use of a neutralizing agent, 152  
102 or membrane filtration, test strains may be added to a test  
103 sample diluted to as maximum as possible extent or to the  
104 final washings of the filter after membrane filtration. If the  
105 product to be examined is used for membrane filtration as it  
106 is, the final washings of the filter after membrane filtration is  
107 inoculated with test strains.

## 108 **6. Addition of culture media**

109 In principle, the type of culture medium specified in a mi-  
110 crobial test cannot be changed. However, depending on the  
111 properties and composition of the product to be examined or  
112 the type of contaminating micro-organisms, it may be more  
113 appropriate to use a culture medium other than that specified.

### 114 **6.1 Non-selective media such as for microbial enumera- 115 tion tests or sterility tests**

116 Perform a growth promotion test to verify the ability to de-  
117 tect micro-organisms, referring to Microbiological Examina-  
118 tion of Non-sterile Products <4.05> and Sterility Test <4.06>,  
119 etc.

120 Some microorganisms require particular components for  
121 growth, while others cannot grow in the presence of particu-  
122 lar components. It should be noted that the number and type  
123 of micro-organisms detected and the appearance of grown  
124 colonies may differ depending on the medium composition,  
125 even using non-selective media.

### 126 **6.2 Selective media for tests for specified microorgan- 127 isms**

128 Perform a growth promotion test to verify the ability to de-  
129 tect micro-organisms, referring to Microbiological Examina-  
130 tion of Non-sterile Products <4.05>, etc. Note that indication  
131 reactions may differ depending on the composition of a cul-  
132 ture medium.

## 133 **7. Incubation time**

134 In the growth promotion tests and the suitability tests,  
135 adopt incubation time specified respectively.

136 However, adopt an appropriate time when it is verified that  
137 the growth of the test organisms can be observed and the  
138 number of organisms can be measured.

139 If the shortest incubation time during the specified incuba-  
140 tion times is used in product testing using the test method  
141 whose suitability is established, the validity of the use of the  
142 shortest incubation time should be verified.

## 143 **8. Counting of the number of colonies in product testing**

144 The counting of the number of colonies by the plate-count  
145 method is performed using an appropriate dilution factor. In  
146 the product testing, the number of colonies of less than 250  
147 CFU per plate for bacteria and yeast, and less than 50 CFU  
148 per plate for moulds as a guide is generally counted. After the  
149 counting, take an arithmetic mean for each medium and mul-  
150 tiple by the dilution factor to calculate the enumeration of vi-  
151 able micro-organisms.