

1 Reference NMR Spectrum Based on ^1H Spin 2 Parameters and Application to Reagents in 3 the Japanese Pharmacopoeia <G5-9-190>

4 (^1H スピン情報に基づいた参照 NMR スペクトルと日本
5 薬局方試薬への応用<G5-9-190>)

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7 For quantitative or qualitative tests of drug products, “JP
8 reference standards” that have appropriate quality for the in-
9 tended purpose have been established. For crude drugs and
10 related drugs, various physicochemical tests targeting marker
11 compounds contained in crude drugs and related drugs have
12 been established and it is desirable to establish JP reference
13 standards of marker compounds for these tests. However, as
14 described in Quantitative Analytical Technique Utilizing Nu-
15 clear Magnetic Resonance (NMR) Spectroscopy and its Ap-
16 plication to Reagents in Japanese Pharmacopoeia <G5-5-170>,
17 marker compounds of crude drugs and related drugs are de-
18 rived from natural resources, and there are various difficul-
19 ties in establishing JP reference standards.

20 In many cases, the Japanese Pharmacopoeia sets the spec-
21 ifications of a marker compound available as a reagent at that
22 time in Reagents, Test Solutions <9.41> of the Japanese Phar-
23 macopoeia, and the reagent is defined as the reference mate-
24 rial for analysis. Since the reagent may contain related sub-
25 stances of the marker compound, purity tests are established
26 in the specifications of the reagent to detect impurities in ad-
27 dition to the identification test. Although chromatographic
28 methods are specified as the purity test in many cases, it is
29 sometimes difficult to separate impurities like diastereomers
30 from marker compounds. On the other hand, Nuclear Mag-
31 netic Resonance Spectroscopy <2.21> is a useful spectro-
32 scopic technique in such discrimination. When ^1H NMR
33 measurement is performed in a superconducting magnetic
34 field, as generally performed at present, even in the case
35 where an impurity having a very similar structure such as a
36 diastereomer is contained, a signal derived from the ^1H nu-
37 cleus of the impurity is observed as a clearly different signal
38 in the measured spectrum, mainly in a part with different
39 structures between the marker compound and the impurity
40 contained in the reagent. Therefore, if ^1H NMR is specified
41 in identification tests, the presence of impurities in the re-
42 agent can be easily confirmed from the obtained spectrum.

43 Originally, ^1H spin parameters such as chemical shifts and
44 spin-spin coupling constants observed from a ^1H NMR spec-
45 trum are specific to a compound and independent of strength
46 of the magnetic field of an NMR apparatus. In addition, ac-
47 curate ^1H spin parameters of a marker compound contained
48 in the reagent can be obtained based on the ^1H NMR spec-
49 trum of the reagent with extremely high purity because soft-
50 ware products for iterative calculation to obtain accurate ^1H
51 spin parameters from measured ^1H NMR spectra can be

52 utilized¹⁾. Moreover, there are software products that can gen-
53 erate the spectrum from the ^1H spin parameters correspond-
54 ing to the magnetic field strength of each measurement¹⁾, so
55 that the reference NMR spectrum corresponding to any mag-
56 netic field strength for identification tests can be easily gen-
57 erated. Therefore, if accurate ^1H spin parameters are speci-
58 fied in the Japanese Pharmacopoeia, identification tests can
59 be performed by direct comparison of spectra in the same
60 manner as the infrared reference spectrum. In addition, de-
61 tection of the distortion and unnecessary signals caused by
62 the existence of structurally similar impurities by comparison
63 of spectra can be used as a purity test. Details of the ^1H spin
64 parameters and precautions to be taken in the identification
65 test using the reference NMR spectrum are described below.

66 1. ^1H spin parameters

67 ^1H spin parameters are generally obtained by measuring
68 the position and multiplicity (peak pattern) of a signal ob-
69 tained directly from a spectrum, and these can be specified as
70 a chemical shift and a spin-spin coupling constant, respec-
71 tively. It is desirable to know the ^1H spin parameters for all
72 protons that are contained in the substance. However, even in
73 the case of a substance with very high purity, the signals may
74 overlap in a limited observation range, resulting in a very
75 complex peak pattern. Therefore, these signals are often de-
76 scribed as multiplet signals, and an accurate reference NMR
77 spectrum cannot be generated because a spectral analysis by
78 observation does not provide accurate ^1H spin parameters.

79 In order to obtain accurate ^1H spin parameters from a spec-
80 trum that consists of such a complex peak pattern, it is effec-
81 tive to use an iterative calculation method. Software products
82 that generate spectra based on ^1H spin parameters have al-
83 ready been available, and software products equipped with
84 an additional function for iterative calculation have become
85 easy to use. In this method, total-line-shape fitting is per-
86 formed each time until iterative calculation converges to ob-
87 tain the ^1H spin parameters (chemical shift and spin-spin cou-
88 pling constant) from the observed spectrum of the substance.
89 When iterative calculation converges and the calculated
90 spectrum agrees with the observed spectrum, the ^1H spin pa-
91 rameters obtained in that time can be regarded as being ex-
92 tremely close to the exact solution. By specifying this ^1H spin
93 parameters in the Japanese Pharmacopoeia, the reference
94 NMR spectrum can be used for the identification test.

95 2. Identification using reference NMR spectrum

96 The NMR spectrum of the sample is obtained by measur-
97 ing ^1H NMR under the specified conditions. When accurate
98 ^1H spin parameters are available for the substance to be ex-
99 amined, a reference NMR spectrum in the same magnetic
100 field as the sample measured above can be generated via the
101 software. When the chemical shift, multiplicity, and relative
102 intensity of each signal in the NMR spectrum of the sample

103 are congruent to those in the reference NMR spectrum, the
 104 sample is judged to meet the identification test. Following
 105 points should be considered for confirming the congruence
 106 of the NMR spectrum of the sample and the reference NMR
 107 spectrum.

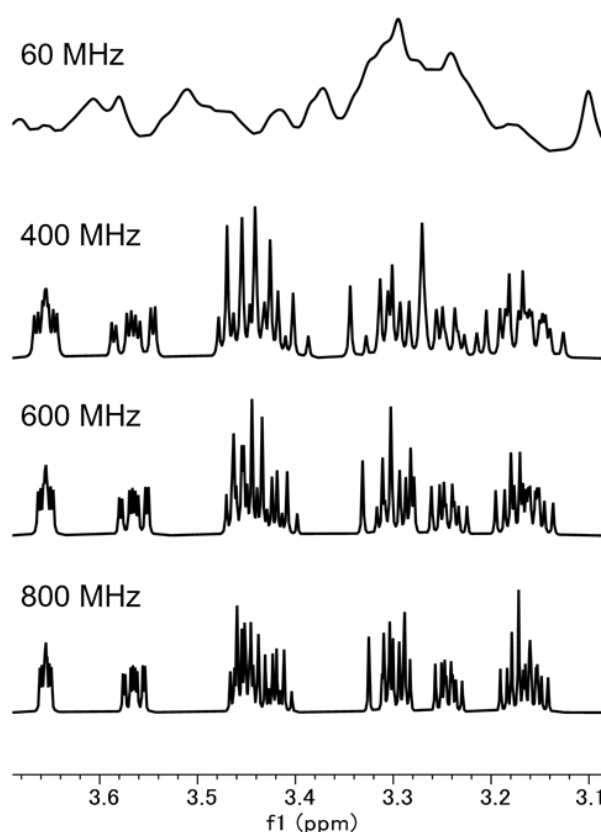
108 2.1. Line width of a signal

109 Fig. 1 shows partial reference NMR spectra of hesperidin
 110 at the ^1H resonance frequency of 60, 400, 600 and 800 MHz.
 111 It can be seen that the peak patterns of the spectra are differ-
 112 ent depending on strength of the magnetic field. Although
 113 these peak patterns are specific to hesperidin in each mag-
 114 netic field strength, it should be noted that the peak pattern
 115 changes in the observation region where the peaks are close
 116 to each other due to the change in the line width of the signal.
 117 In generating the reference NMR spectra shown in Fig. 1, the
 118 line width of the signal has been standardized to 1 Hz.

119 In the spectrum of a sample, the line width of its signals
 120 are changed by adjusting the shim and/or the resolution of the
 121 apparatus. Therefore, when confirming the congruence of the
 122 NMR spectrum of the sample and the reference NMR spec-
 123 trum of hesperidin, it is desirable to match the line width of a
 124 signal of the reference NMR spectrum with that of the sample
 125 or to apply a broadening factor of about 2 Hz to both spectra.

126 2.2. Signals derived from deuterated solvents and water

127 In the NMR spectrum of a sample, signals derived from
 128 the deuterated solvent used to dissolve the sample and from
 129 water, as well as signals from a reference compound for
 130 chemical shifts when it is used, can be observed. Note that in
 131 the case where these signals overlap with those of the marker
 132 compound in the sample, this region should not be used to
 133 confirm the congruence of the spectrum to the reference
 134 NMR spectrum. Since signals derived from slight impurities
 135 in the deuterated solvent may be observed, it is desirable to
 136 measure a blank containing only the deuterated solvent in ad-
 137 vance to confirm the range and intensity of the observed sig-
 138 nals of the impurities. In addition, the signal derived from
 139 water tends to increase in the case of a highly hygroscopic
 140 deuterated solvent which has been stored for a while after the
 141 container is opened. Therefore, it is desirable to use freshly
 142 opened or ampule type deuterated solvents.



143
 144 **Fig. 1** Reference NMR spectra of hesperidin in deuterated di-
 145 methyl sulfoxide at ^1H resonance frequencies of 60, 400, 600
 146 and 800 MHz

147 3. References

- 148 1) P.S. Achanta, *et al.*, J. Pharm. Biomed. Anal., 192,
 149 113601 (2021).

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